



**National B Virus Resource Center
Viral Immunology Center
Georgia State University
161 Jesse Hill Jr. Drive
Atlanta, Georgia 30303**



B VIRUS EXPOSURE MINI-PROTOCOL

The following is a checklist for recommended samples to be submitted for herpes B virus testing. Please familiarize yourself and your staff with this checklist and please refer to this when submitting specimens to our laboratory. Thank you.

BASELINE/DAY OF INJURY SAMPLES

- 1. Human baseline serum (0.5 - 2.0 ml) collected as close as possible to the time of injury.
- 2. Primate baseline serum (0.5 - 2.0 ml) collected as close as possible to the time of injury.
- 3. Human virology swab samples of the wound site or exposed area as applicable. This specimen should be collected as soon as possible after the injury, after the site has been disinfected.
- 4. Primate virology swab samples of the buccal cavity, right eye, left eye, and genitalia. Please use one swab per site and send in separate media tubes. These specimens must be collected as close as possible to the time of injury, as specimens collected later may not accurately reflect the monkey's status at the time of injury.

FOLLOW-UP/14 - 21 DAY POST INJURY SAMPLES

- 1. Human follow-up serum (0.5 - 2.0 ml).
- 2. Primate follow-up serum (0.5 - 2.0 ml).

IMPORTANT COMMENTS

1. Please refer to the document *Recommendations on Sample Collection, Storage, & Shipment* for complete instructions for specimen collection, handling, and shipment.
2. **Because we test paired specimens, the failure to submit a baseline or follow-up serum sample will result in insufficient specimens for complete testing, potentially resulting in unreliable diagnosis.**
3. Fill out the submission form completely and correctly. Please verify that all information is **identical** to the specimen labels.
4. Label all specimens clearly with the permanent name or ID, date of collection, virology swab collection site, and/or tissue source. Failure to correctly label specimens may result in incomplete results. Mislabeled or unlabeled specimens may not be tested.
5. Do not label specimen tubes with extra information that is not indicated above. Cage #'s, study #'s, experiment #'s, investigator's name, etc. are unnecessary and confusing when trying to identify the sample.
6. Be sure all whole blood samples (if submitted for serum antibody testing) are spun and separated. Remove the serum and transfer to a properly labeled plastic tube for shipment to our laboratory.
7. On occasion, it will not be possible to provide our laboratory with the requested specimens because the associated monkey is unidentifiable, was euthanized, the injury was reported late, etc. If for any reason you are unable to collect the appropriate specimens, please note that information on the submission form.

(OVER)

Address packages to:

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Georgia State University
161 Jesse Hill Jr. Drive
Atlanta, Georgia 30303**

Address correspondence to:

**National B Virus Resource Center
Viral Immunology Center
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Emergency Phone Numbers:

National B Virus Resource Center:

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Emergency

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Primary Clinical Consultants

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PRE-SHIPMENT CHECK-LIST

Before shipping your samples to our laboratory, please make sure you have:

(reference instructions, *Recommendations on Sample Collection, Storage, & Shipment*)

- contacted our laboratory to alert us of your shipment.
- used appropriate primary and secondary shipping containers with adequate absorbent material and used the proper labels on the outside of the containers (reference *Federal Register* 42 CFR Part 72).
- packed with at least 5 pounds of appropriate coolant.
- used appropriate delivery address.
- not used glass specimen tubes.
- provided a contact name and phone number in case of emergency.
- marked the package and courier form for "SATURDAY DELIVERY" if shipping on a Friday.
- that the paperwork is properly filled out and that the specimen tubes are labeled to match the paperwork.